

Assembling and Testing Physically Based Models of Observed Complex Systems

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This talk is about the general principles of learning about model states and parameters from laboratory or field data. We want to transfer the information in the (noisy) data to a model (with model errors) of the physical or biological dynamical processes producing the measurements.

When unknown model parameters and unobserved state variables are estimated using the data, we can validate the model through prediction when new forcing is presented.

Using the same methods one can: design experiments; determine how many measurements are required; retrieve more information from existing measurements.

Examples will be discussed for

**biophysical modeling of neurons and functional networks
neuromorphic engineering---neurons and functional networks on a chip**

Some application areas for Data Assimilation:

Genetic regulatory networks

signal transduction pathways

systems biology; synthetic biology; Immunology

biophysical modeling of neurons and functional networks

neutrino astrophysics

coastal flows and transport of toxic constituents after storms

electrical and chemical engineering

identifying oil and gas reservoirs

hydrological models of streams and lakes

neuromorphic engineering---neurons and functional networks on a chip

numerical weather prediction

Neuron Model

$$C \frac{dV(t)}{dt} = g_{Na} m(t)^3 h(t) (E_{Na} - V(t)) + g_K n(t)^4 (E_K - V(t)) + g_L (E_L - V(t)) + I_{applied}(t)$$

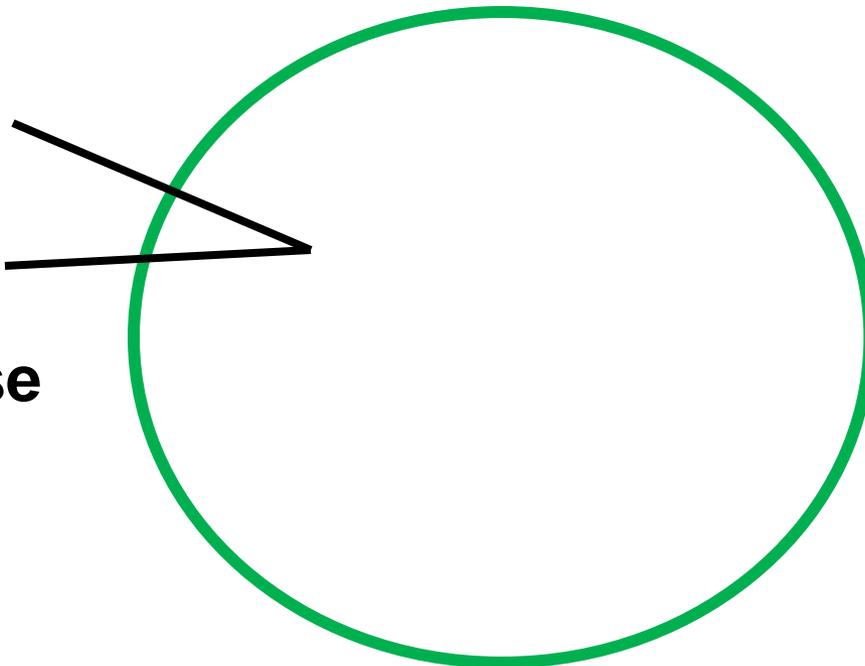
$$\frac{da(t)}{dt} = \frac{a_\infty(V(t) - a(t))}{\tau_x(V(t))} \quad a(t) = \{m(t), h(t), n(t)\}$$

$$a_\infty(V) = \frac{1}{2} \left(1 + \tanh\left(\frac{V - V_{x}}{dV_x}\right) \right)$$

$$\tau_x(V) = t_{1x} + t_{2x} \left(1 - \tanh^2\left(\frac{V - V_{xt}}{dV_{xt}}\right) \right)$$

**Inject
current
 $I_{applied}(t)$**

**Measure
Response
Voltage
 $V(t)$**



D = 4 L = 1 p = 20

**Measure $V(t)$ with
selected $I_{applied}(t)$**

**Evaluate all
parameters and all
unobserved state
variables $a(t)$**

This is the challenge:

Using laboratory experiments on individual neurons and on collections of neurons. **Build biophysically based models of functional neural networks which matches experiments and predicts the response to new stimuli.**

Our strategy is this:

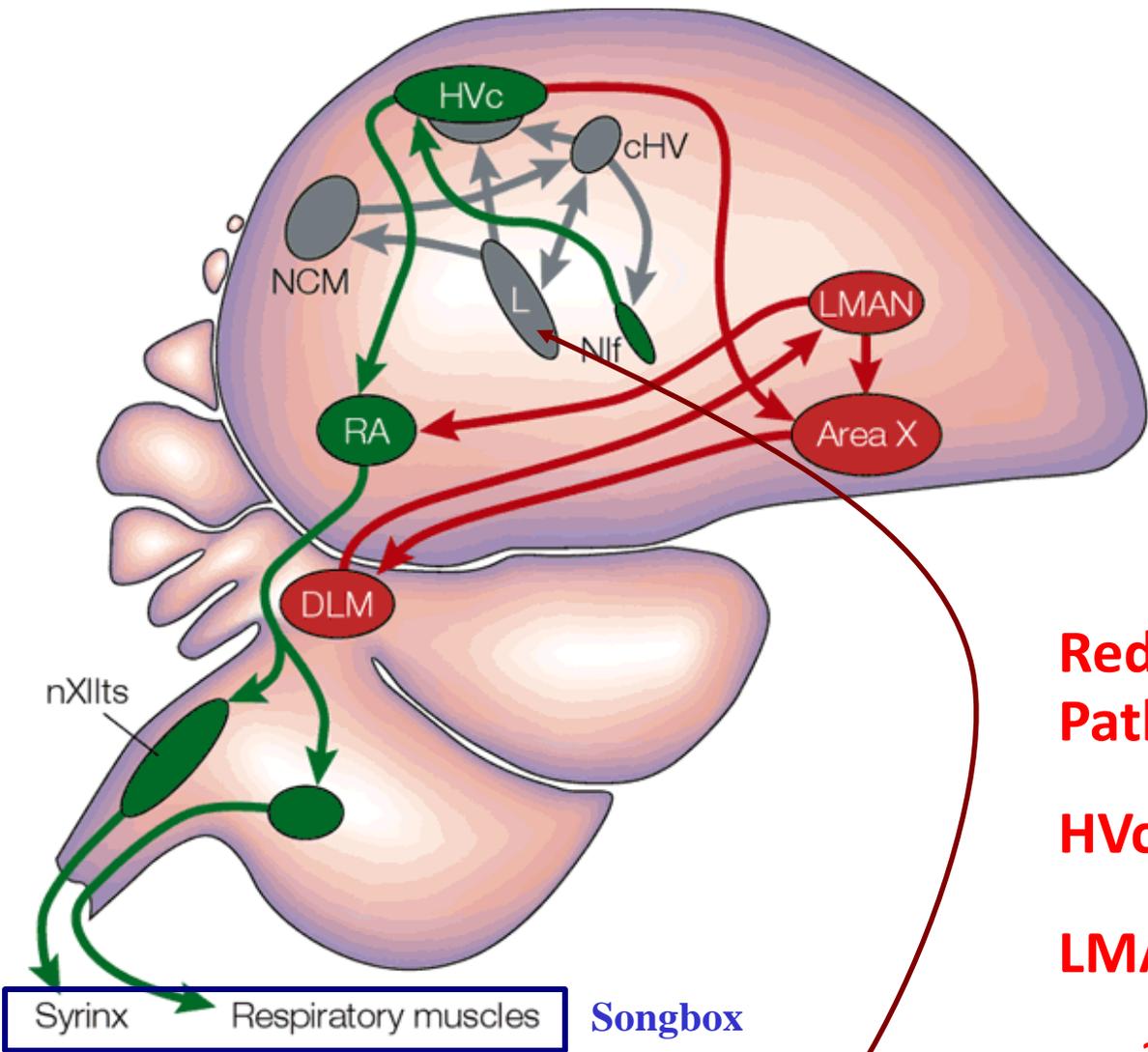
- create a model of the functional network of interest—e.g. song production network for song birds and, of course, the individual neurons in the network---what is a sensible model—we use Hodgkin-Huxley models.
- using the model itself, design experiments that stimulate all degrees of freedom of the neuron/network and measure enough quantities at each observation time---these are numerical simulations.
- Use the model along with data, voltage across membranes, and perhaps other measurements, to determine the unknown parameters in the model and the unobserved state variables in the model.
- “validate the neuron model”—via prediction—These validated neurons can be used in network construction.

One mainstream view of network modeling and operation is that details do not matter but some form of network “organization” or structure determines network operations.

Our use of data assimilation to design experiments, test and validate models of cells and systems points to advantages of other directions.

Why would we want the kind of detail of neural or cellular processes accurate modeling and careful data assimilation provides?

- **use models of nerve cells (neurons) to compare healthy and diseased cells to provide biophysical targets for therapies**
- **use detailed models of regulatory networks for genetic action to design interventions**
- **use detailed, verified models of functional network connectivity and nodal performance to engineer functions into high-performance electronics---e.g. sequence generation and recognition with human accuracy but machine performance**



Green: Motor Pathway

HVC → RA →

Respiration/Syrinx

Song Production

Red: Anterior Forebrain Pathway (AFP)

HVC → Area X → DLM →

LMAN → Area X

and HVC

Control and Song

Maintenance

Auditory Feedback

Songbox

Syrinx Respiratory muscles

Isolated Neurons from the Avian Song System

On each neuron many different $I_{\text{applied}}(t)$ measurements in time “epochs” of 2-6 seconds

Membrane Voltage Observed

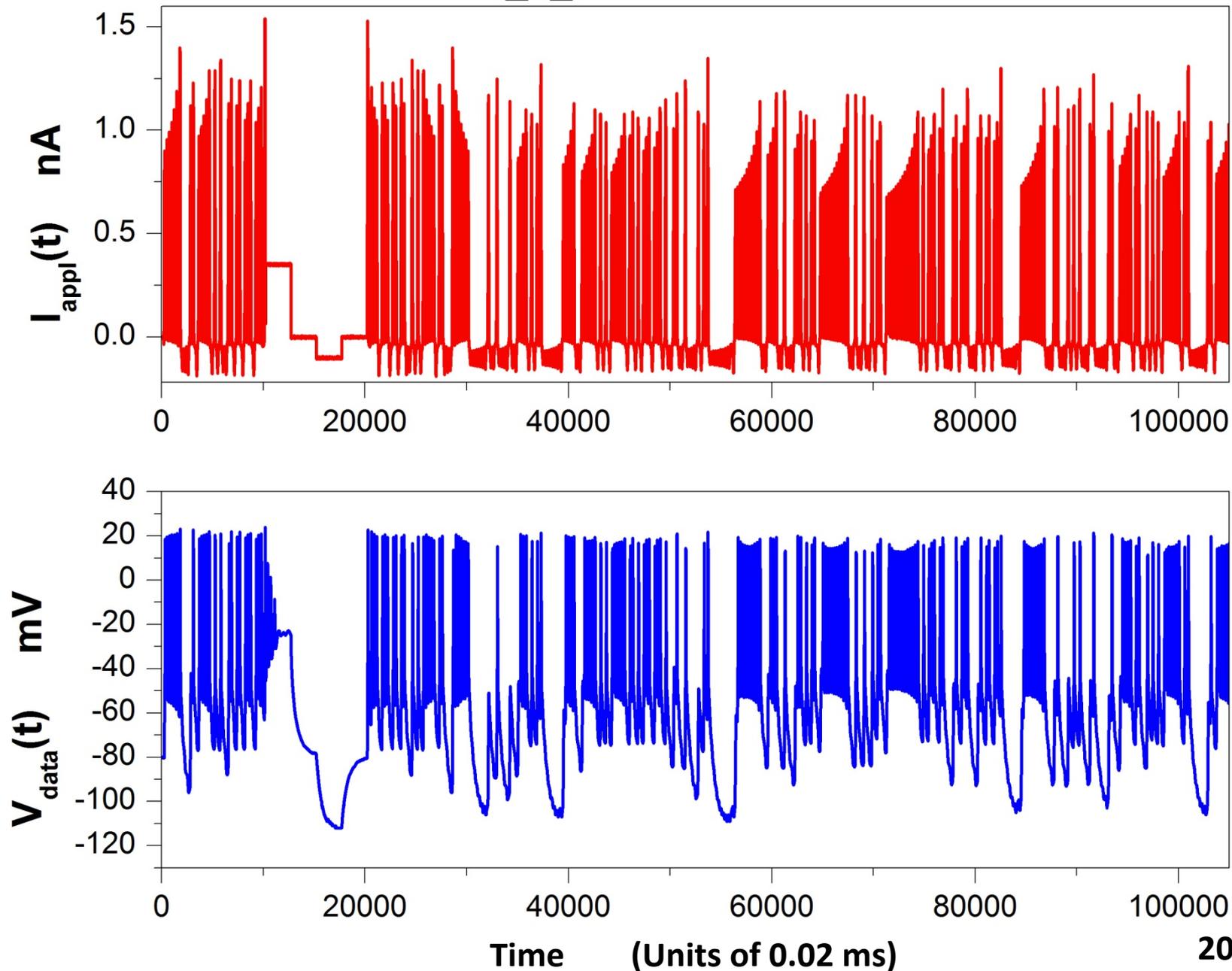
Sampling time 0.02 ms (50 kHz), 500-1500 ms of observations

Use all this to estimate the unknown parameters in the neuron and unmeasured state variables, then predict the response of the neuron to new stimuli (forcing). **Model Validation**

**Neurobiological Laboratory Experiments
Margoliash Laboratory, UChicago**

0512_1_3 Neuron Data

Why this $I_{\text{appl}}(t)$?



Data Assimilation:

Transfer of Information from Measurements to a Model of the Observations

We have

noisy measurements $y_k(t)$; $k = 1, \dots, L$; and

errors in the model $x_a(t+\Delta t) = f_a(x(t))$; $a = 1, \dots, D \gg L$,

uncertain initial conditions at the starting time t_0 .

We make measurements in a time window $[t_0, T]$. We wish to incorporate the information in these measurements at $t_0, t_1, \dots, t_N = T$ into our statistical estimate of the complete state of the model at T and into our statistical estimate of the model parameters.

Statistical data assimilation is communication of information from measurements (transmitter) to a dynamical model (receiver).

At the end of an observation window $[t_0, T]$ we want the conditional probability distribution of the state $x(T)$ of the system

$$\mathbf{\underline{P}(x(T)|\text{observations})}$$

given measurements during the window.

We then want to predict the future conditional probability distribution $P(x(t > T))$ for new forcing of the system.

1st
observation
 $y(t_1)$

2nd
observation
 $y(t_2)$

$y(t_n)$

$y(t_{n+1})$

$P(x(t_1)|x(t_0))$

$P(x(t_2)|x(t_1))$

$P(x(t_{n+1})|x(t_n))$

$P(x(t_0))$ $P(y(t_1)|x(t_0))$ $P(y(t_2)|x(t_1),x(t_0))$ $P(y(t_n)|x(t_0),\dots,x(t_{n-1}))$

Time→

t_0

t_1

t_2

t_n

t_{n+1}

$t_N = T$

Start here

Conditional probability of states $X(N)$ given observations $Y(N)$:
 $P(X(N)|Y(N))$

$$X(N) = \{x(t_0), x(t_1), \dots, x(t_N)\} \quad Y(N) = \{y(t_0), y(t_1), \dots, y(t_N)\}$$

$$P(X(N)|Y(N)) = \prod_{k=0}^N P(y(k) | X(k), Y(k-1)) \prod_{k=0}^{N-1} P(x(n+1) | x(n)) P(x(0))$$

Information Transfer

Dynamics of System

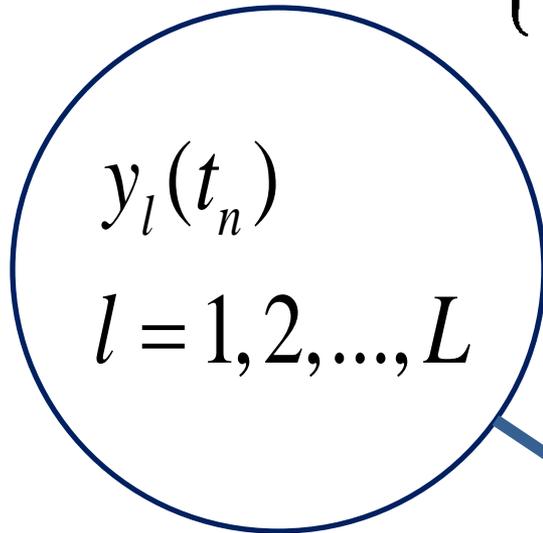
$$= e^{-A_0(X)}$$

Expected value of functions $G(X)$ on the path $X(N)$,

$$\langle G(X) \rangle = \frac{\int dX e^{-A_0(X)} G(X)}{\int dX e^{-A_0(X)}}$$

e.g. $G(X) = X$; $X - \langle X \rangle$; other moments

$$t = \{t_0, t_1, \dots, t_n, \dots, t_N = T\}$$



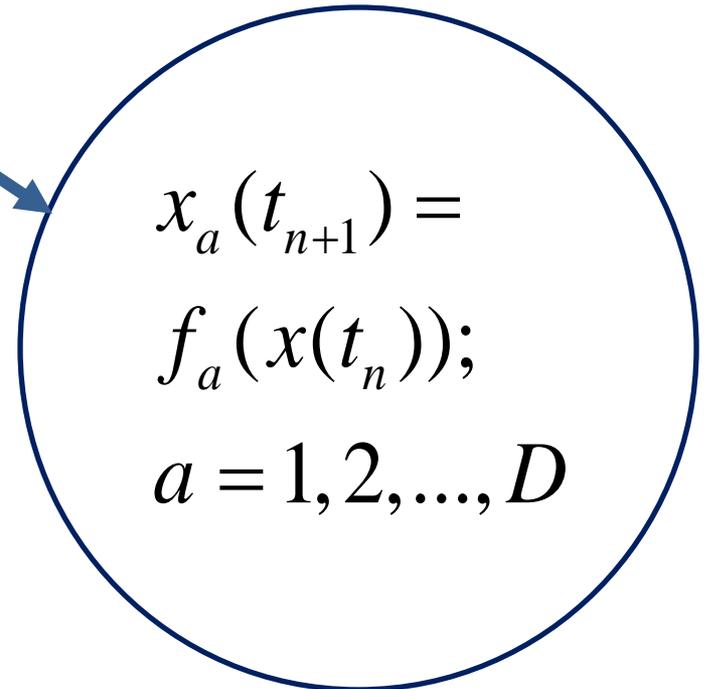
Data source: **Transmitter**

$$L \leq D$$

$$y_l(n) = h_l(x(n))$$

**Generalized synchronization of
the transmitter and receiver**

Model: **Receiver**



In neurobiology: We have measurements of voltage across the cell membrane during the observation window

$$Y = \{y(t_0) = V(t_0), y(t_1) = V(t_1), \dots, y(T) = V(T)\} \quad (\text{curr}$$

$L = 1, D = 12, p = 50$ Maximal conductances g_{Na}, g_{SK}, \dots

We have a model (current conservation)

$$C dV(t)/dt = I_{Na}(t) + I_K(t) + I_{Ca}(t) + I_{SK}(t) + \dots + I_{stimulus}(t)$$

$$\frac{dx(t)}{dt} = F(x(t))$$

Measurements have errors, model has errors

$$E[G(X)|Y] = \langle G(X) \rangle = \frac{\int dX e^{-A_0(X)} G(X)}{\int dX e^{-A_0(X)}}$$

Path is through state variable+parameter space

$X = \{x(N), x(N-1), \dots, x(0)\}$ including parameters

Everything hinges on the structure of $A_0(X)$ in path space

Two methods for evaluating these high dimensional integrals:

(1) Laplace's method (1774); seek minima of $A_0(X)$ --there are multiple minima

$$\frac{\partial A_0(X)}{\partial X_\alpha} = 0 \quad \text{and} \quad \frac{\partial^2 A_0(X)}{\partial X_\alpha \partial X_\beta} > 0$$

(2) Monte Carlo searches of $\exp[-A_0(X)]$ —distribution in path space.

First method seeks minima of $A_0(X)$

Second method samples near these minima.

We focus on the Laplace method. How can we find the minima X^q $q=0,1,\dots$

$$\left. \frac{\partial A_0(X)}{\partial X_\alpha} \right|_{X^q} = 0 \quad q=0,1,\dots \quad A_0(X^0) < A(X^{q \neq 0})$$

Which minimum gives the biggest contribution to the integral ?
Smallest action $A_0(X^0)$ from path X^0

Everything rests on the structure of $A_0(X)$ in path space.

Action for State and Parameter Estimation

Information Transfer

$$A_0(X) = - \sum_{n=0}^N \log \{ CMI(y(n) | X(n), Y(n-1)) \}$$

$$- \sum_{n=0}^{N-1} \log \{ P(x(n+1) | x(n)) \} - \log \{ P(x(0)) \}$$

Dynamics

Initial Condition

$A_0(X)$ is nonlinear in X and has multiple minima. Location and number of these minima depend on the model and on the number of measurements at each observation time in $[t_0, t_N]$.

Standard model, Gaussian Error Action

If observations have Gaussian noise and models have Gaussian errors, action is

$$A_{\text{GEA}}(X) = \sum_{n=0}^N \sum_{l=1}^L \frac{R_m(n, l)}{2} (x_l(n) - y_l(n))^2 +$$

$$\sum_{n=0}^{N-1} \sum_{a=1}^D \frac{R_f(a)}{2} (x_a(n+1) - f_a(x(n)))^2$$

This is not Gaussian, if $f(x)$ is nonlinear. Finding the minima at any R_m and R_f is not hard (IPOPT), but finding the path with the smallest action—a challenge

Looking in continuous time shows the challenge clearly:

$$\begin{aligned}
 A_{GEA}(x(t), \dot{x}(t)) &= \int_{t_0}^{t_f} dt L(x(t), \dot{x}(t)) \\
 &= \int_{t_0}^{t_f} dt \left[\sum_{l=1}^L \frac{R_m(t)}{2} (x_l(t) - y_l(t))^2 + \sum_{a=1}^D \frac{R_f(a)}{2} (\dot{x}_a(t) - F_a(x(t)))^2 \right]
 \end{aligned}$$

$\delta A_0(X) = 0$ gives the Euler-Lagrange equations:

$$\left(\delta_{ab} \frac{d}{dt} + DF_{ab}(x(t)) \right) \left(\frac{dx_b(t)}{dt} - F_b(x(t)) \right) = \delta_{al} \frac{R_m}{R_f} (x_l(t) - y_l(t))$$

'nudging'

With the boundary conditions

$$p_a(t_0) = p_a(t_f) = 0.$$

$$p_a(t) = \frac{\partial L(x(t), \dot{x}(t))}{\partial \dot{x}_a(t)}$$

$$A_{\text{GEA}}(X) = \sum_{n=0}^N \sum_{l=1}^L \frac{R_m(n,l)}{2} (x_l(n) - y_l(n))^2 + \sum_{n=0}^{N-1} \sum_{a=1}^D \frac{R_f(a)}{2} (x_a(n+1) - f_a(x(n)))^2$$

To determine the paths for the lowest minimum action; find minimum for very small model error value R_f , then move slowly in R_f to a larger value, then an even larger value. We call this **annealing**; distinct from standard simulated annealing.

If $R_f \rightarrow \infty$, model error is 0. We look at the opposite limit $R_f \rightarrow 0$, where model plays no role and dynamical phase space structure is absent.

At $R_f = 0$, minimum is degenerate at $x_l(t) = y_l(t)$; other unmeasured states undetermined. With $R_f = R_{f0}$ very small, choose N_0 initial starting paths with $x_l(t) = y_l(t)$, others chosen from a uniform distribution, this is a set of X_0 for numerical minimization using IPOPT. We call outcomes X_1 .

Use these as initial starting paths with $R_f = \alpha R_{f0}$; $\alpha > 1$, to arrive at N_0 paths X_2 . Increase R_f to $\alpha^2 R_{f0}$, ... and continue using outcome paths as initial guesses for next optimizations, slowly increasing R_f by powers of α .

Plot $A_0(X^q)$ versus $\beta = \log_{\alpha} [R_f / R_{f0}]$. Action level plots.

Simple Model Neuron NaKL

$D+N_p = 4 + 19, L = 1$ Voltage

Twin Experiment on NaKL Neuron

$$C \frac{dV(t)}{dt} = g_{Na} m(t)^3 h(t) (E_{Na} - V(t)) + g_K n(t)^4 (E_K - V(t)) + g_L (E_L - V(t)) + I_{applied}(t)$$

$$\frac{dx(t)}{dt} = \frac{x_\infty(V(t) - x(t))}{\tau_x(V(t))} \quad x(t) = \{m(t), h(t), n(t)\}$$

$$x_\infty(V) = \frac{1}{2} \left(1 + \tanh\left(\frac{V - V_x}{dV_x}\right) \right)$$

$$\tau_x(V) = t_{1x} + t_{2x} \left(1 - \tanh^2\left(\frac{V - V_{xt}}{dV_{xt}}\right) \right)$$

Generate data from NaKL equations

$y(t) = x(t) + \sigma N(0,1)$ noise

D = 4, L = 1

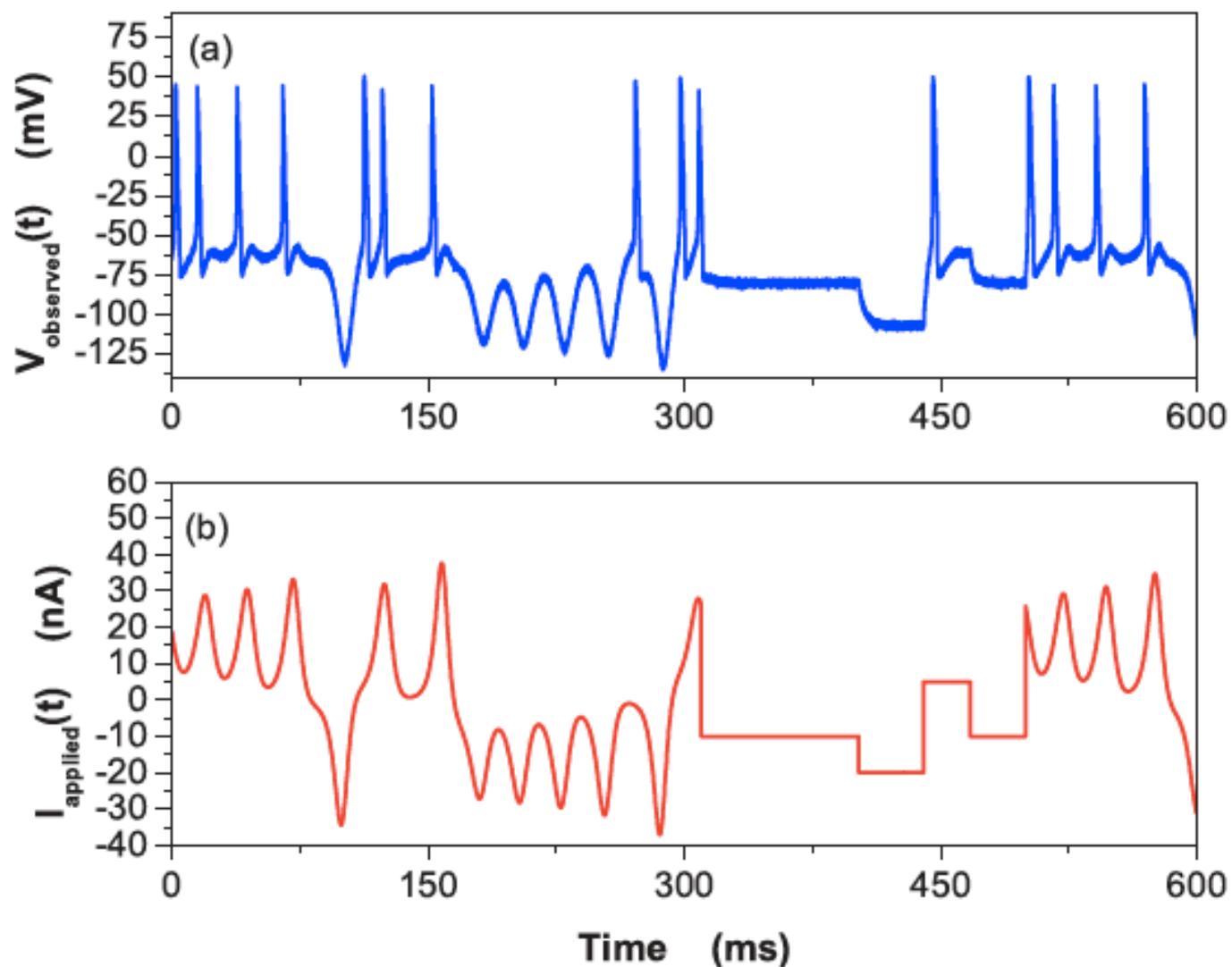


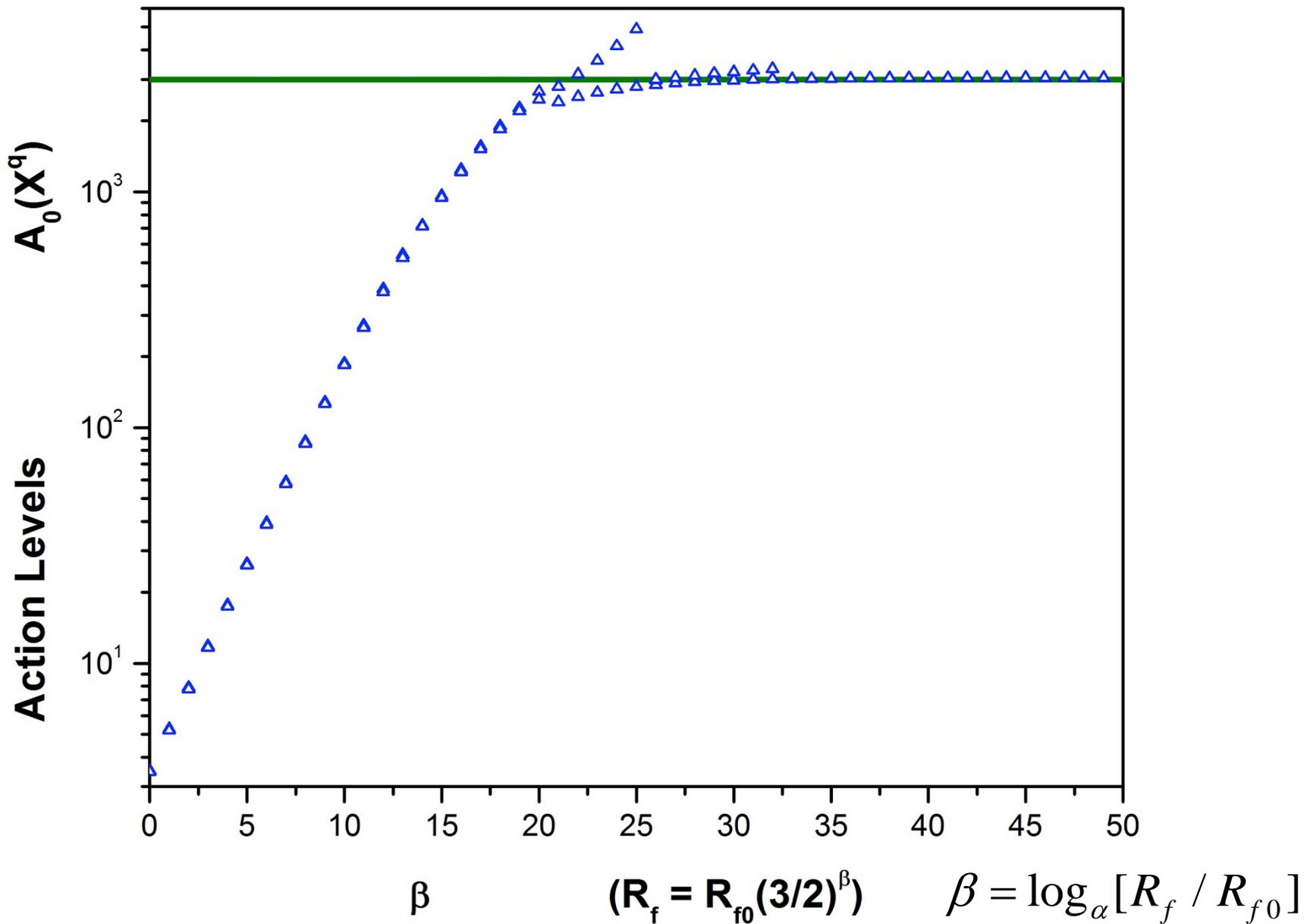
FIG. 8. (Color online) (a) Voltage response of the standard Hodgkin-Huxley neuron model with Na^+ , K^+ , and leak channels, our NaKL model, in response to the applied (injected) stimulus current shown in the (b).

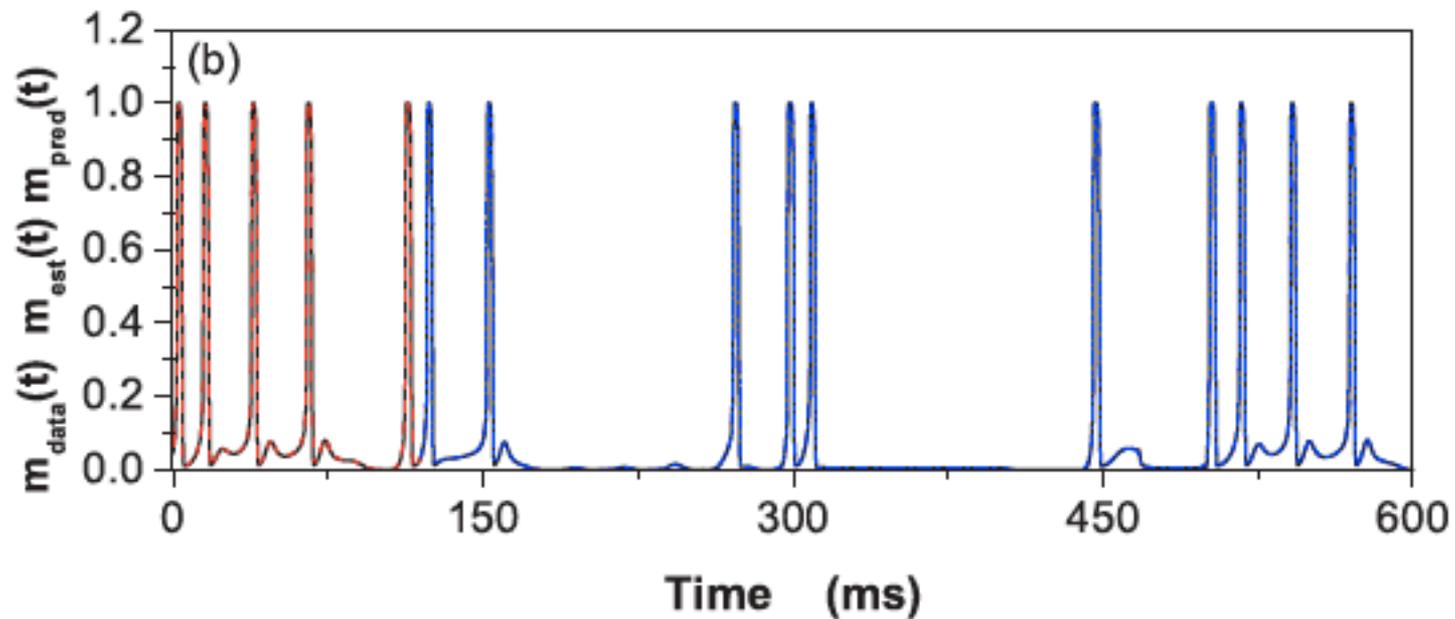
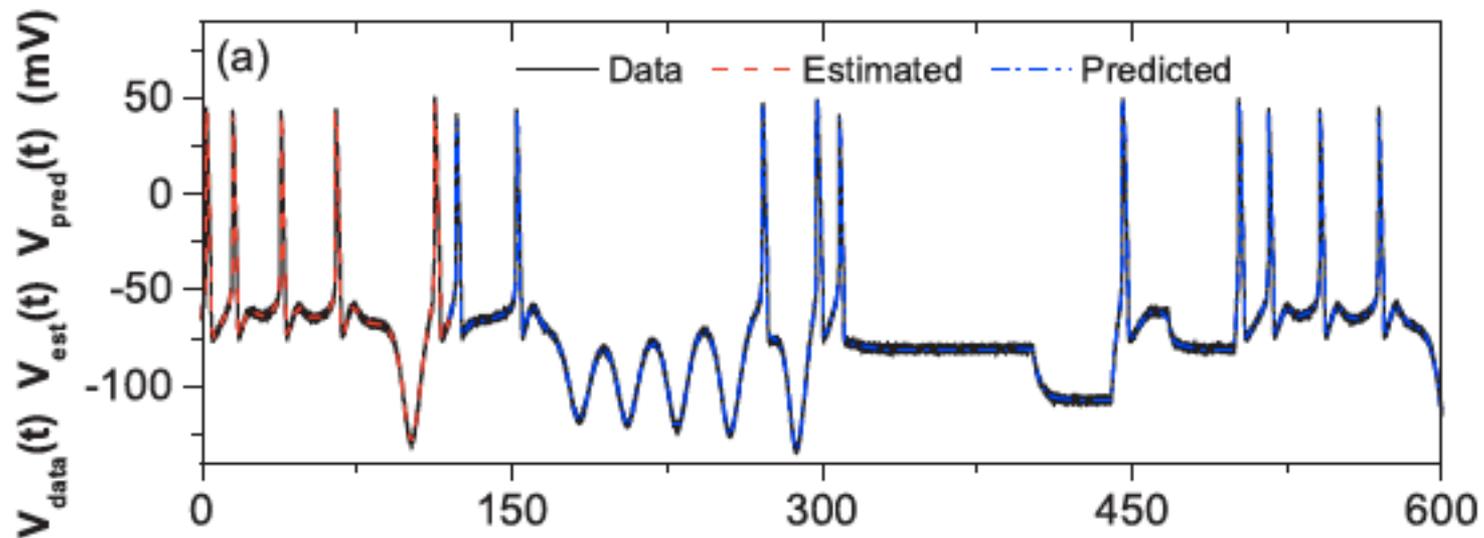
Annealing in Model Error Accuracy

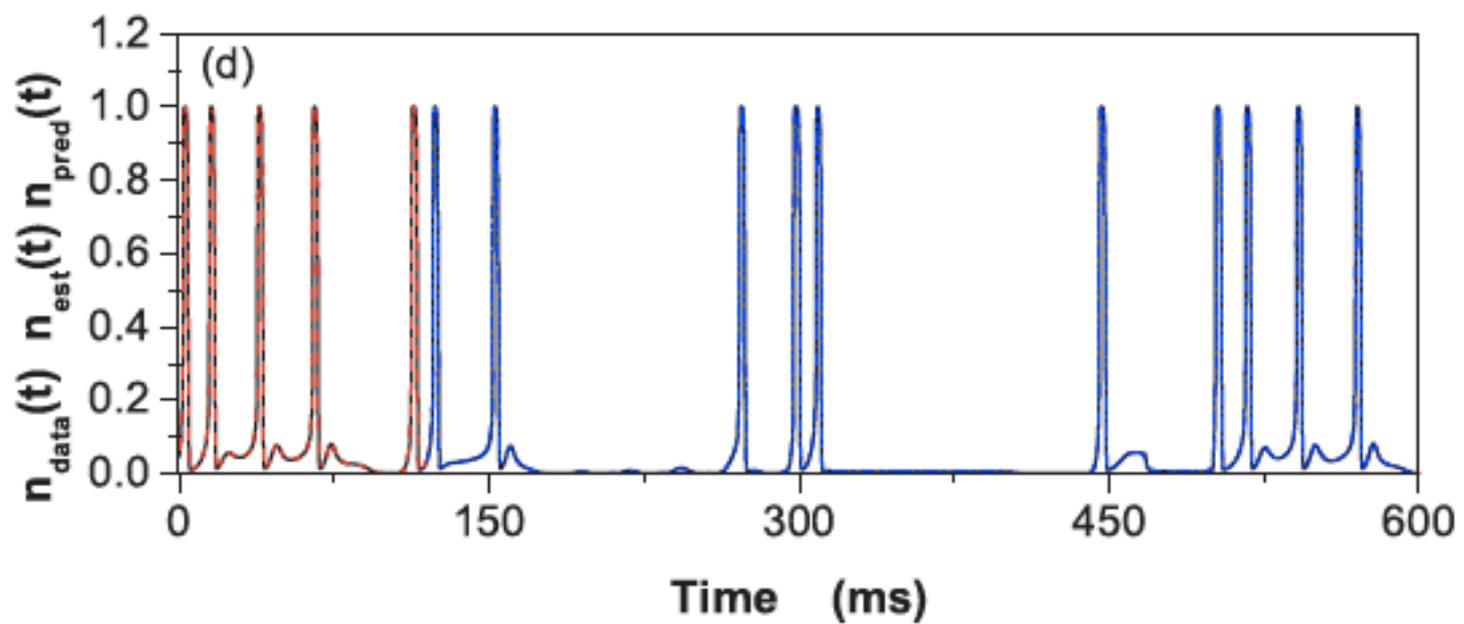
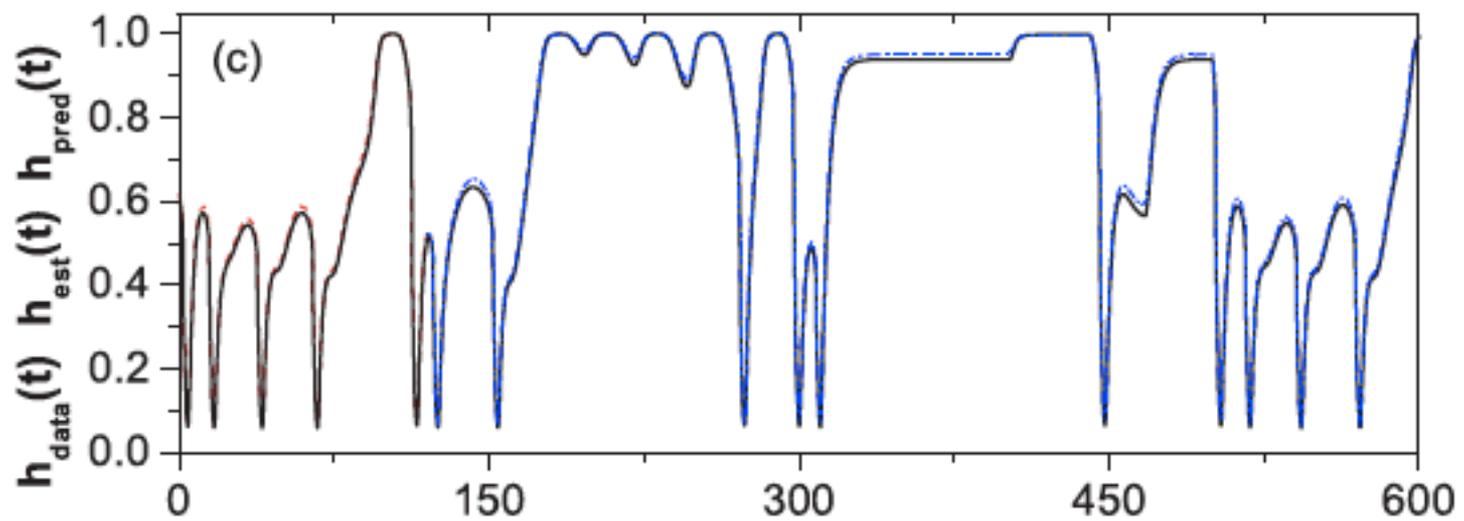
Paths giving minima of the action depend on the number of measurements L .

For the **Standard Model** GEA, when action levels are independent of R_f , the action level is dictated by statistics of measurement error term---a consistency check on the action level evaluations.

NaKL Model Action Level Plot







NaKL Neuron

Twin Experiment

Parameters	Known	Estimated	LB	UB
g_{Na}	120.0	108.4	50.0	200.0
E_{Na}	50.0	49.98	0.0	100.0
g_K	20.0	21.11	5.0	40.0
E_K	-77.0	-77.09	-100.0	-50.0
g_L	0.3	0.3028	0.1	1.0
E_L	-54.0	-54.05	-60.0	-50.0
C	0.8	0.81	0.5	1.5
V_m	-40.0	-40.24	-60.0	-30.0
dV_m	0.0667	0.0669	0.01	0.1
τ_{m0}	0.1	0.0949	0.05	0.25
τ_{m1}	0.4	0.4120	0.1	1.0
V_h	-60.0	-59.43	-70.0	-40.0
dV_h	-0.0667	-0.0702	-0.1	-0.01
τ_{h0}	1.0	1.0321	0.1	5.0
τ_{h1}	7.0	7.76	1.0	15.0
V_n	-55.0	-54.52	-70.0	-40.0
dV_n	0.0333	0.0328	0.01	0.1
τ_{n0}	1.0	1.06	0.1	5.0
τ_{n1}	5.0	4.97	2.0	12.0

Lorenz96 Model $D = 5$

$$\frac{dx_a(t)}{dt} = x_{a-1}(t)(x_{a+1}(t) - x_{a-2}(t)) - x_a(t) - f$$

$$a = 1, 2, \dots, D;$$

$$x_{-1}(t) = x_{D-1}(t); x_0(t) = x_D(t); x_{D+1}(t) = x_1(t).$$

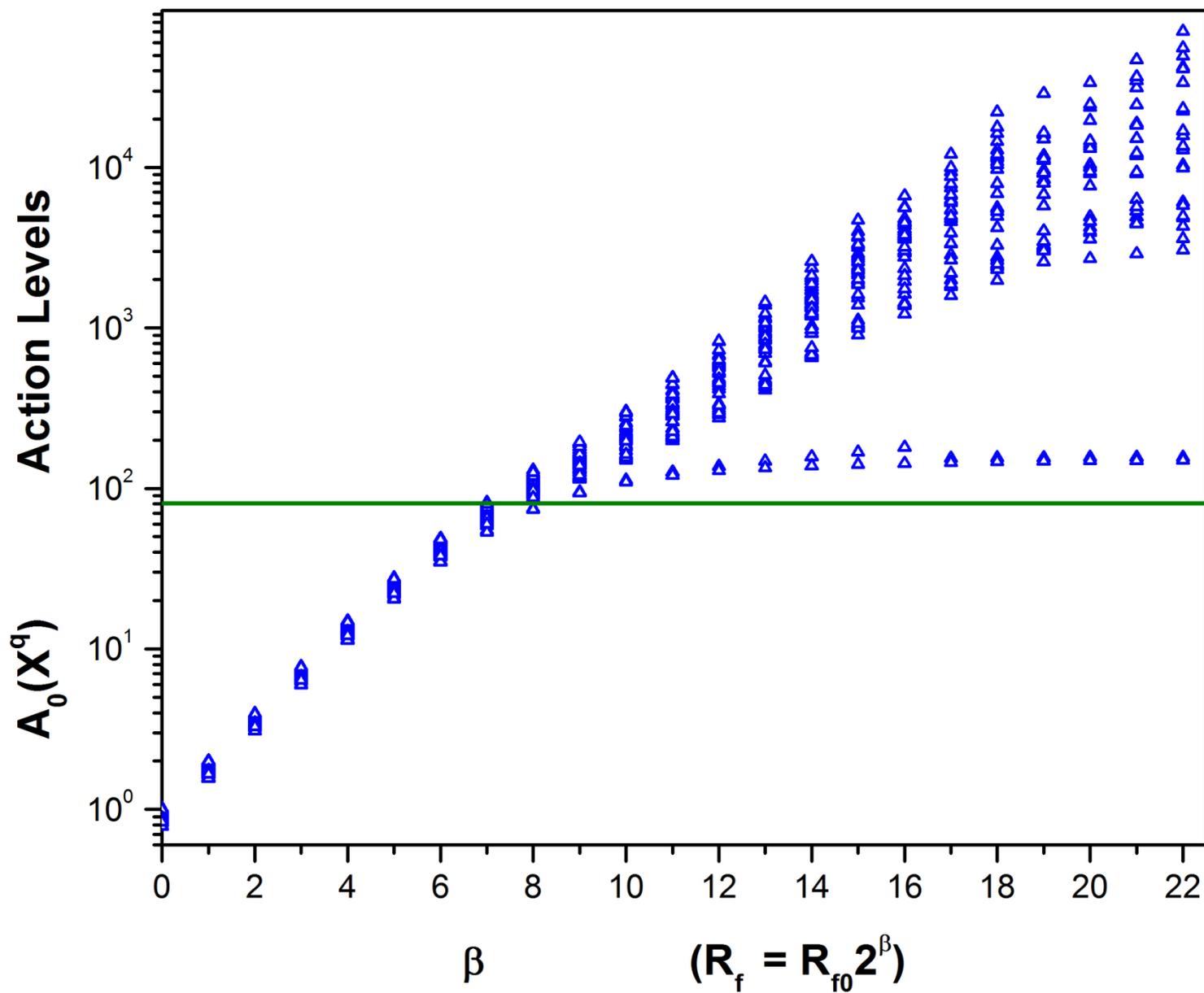
f is a fixed parameter $f = 8.17$. Solutions are chaotic.

‘Twin Experiments’ Use to test methods of data assimilation; use to design experiments.

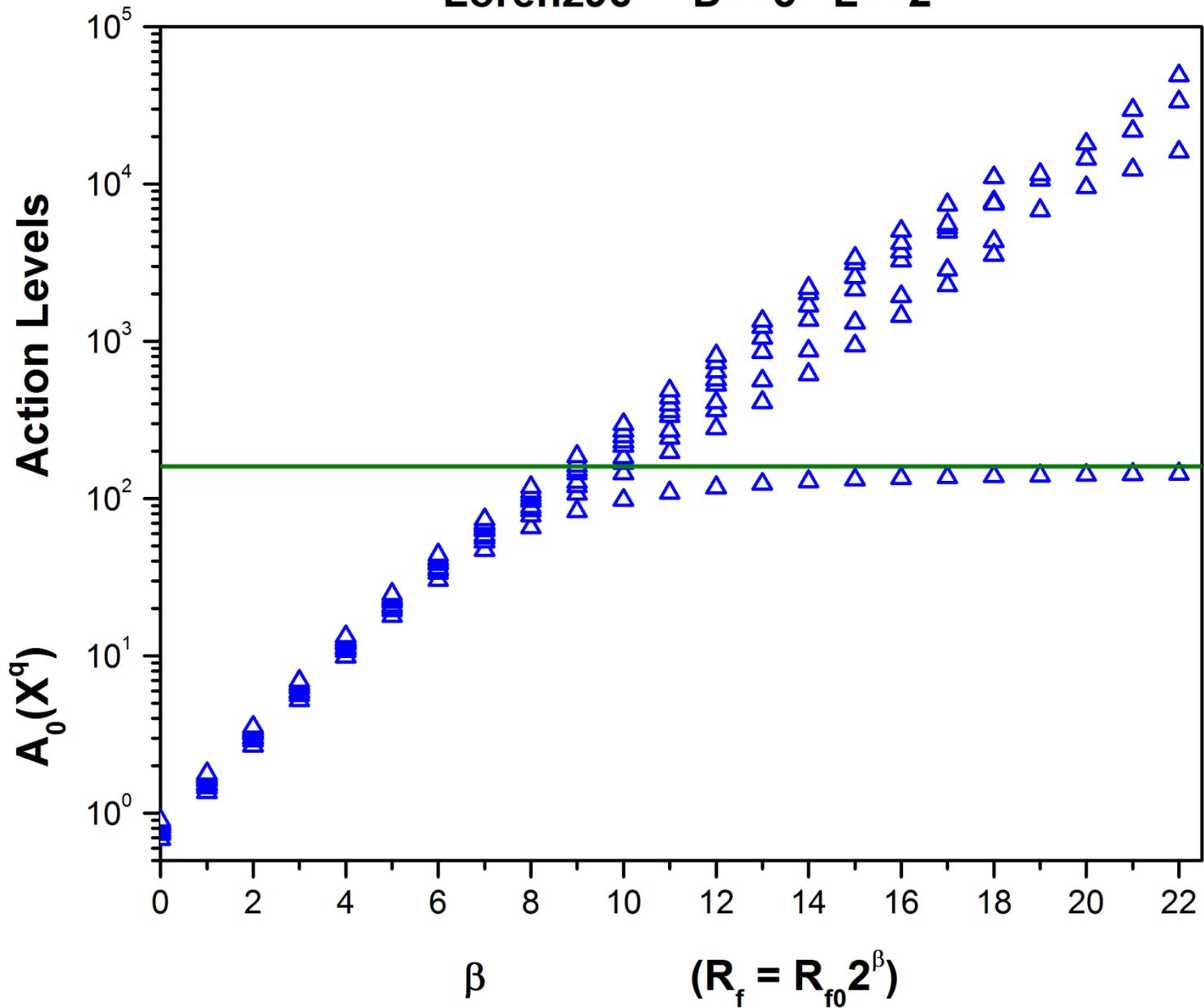
Generate data with known model; add noise to model output; present $l = 1, 2, \dots, L < D$ time series to assimilation procedure.

Lorenz96 Model

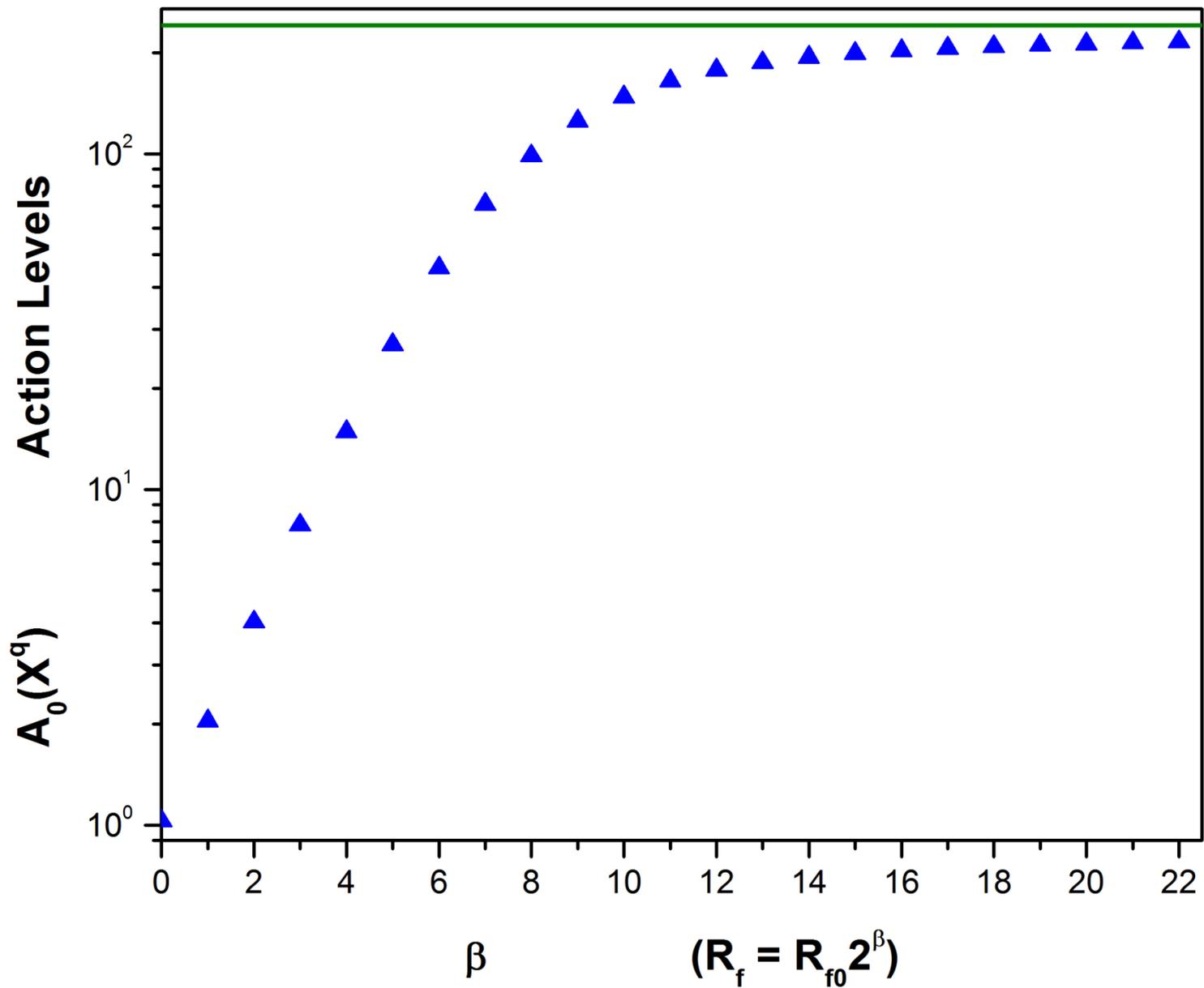
D = 5, L = 1



Lorenz96 D = 5 L = 2



Lorenz96 D = 5 L = 3



Back to Song System Nucleus HVC

Interneurons, $L = 1$ Voltage

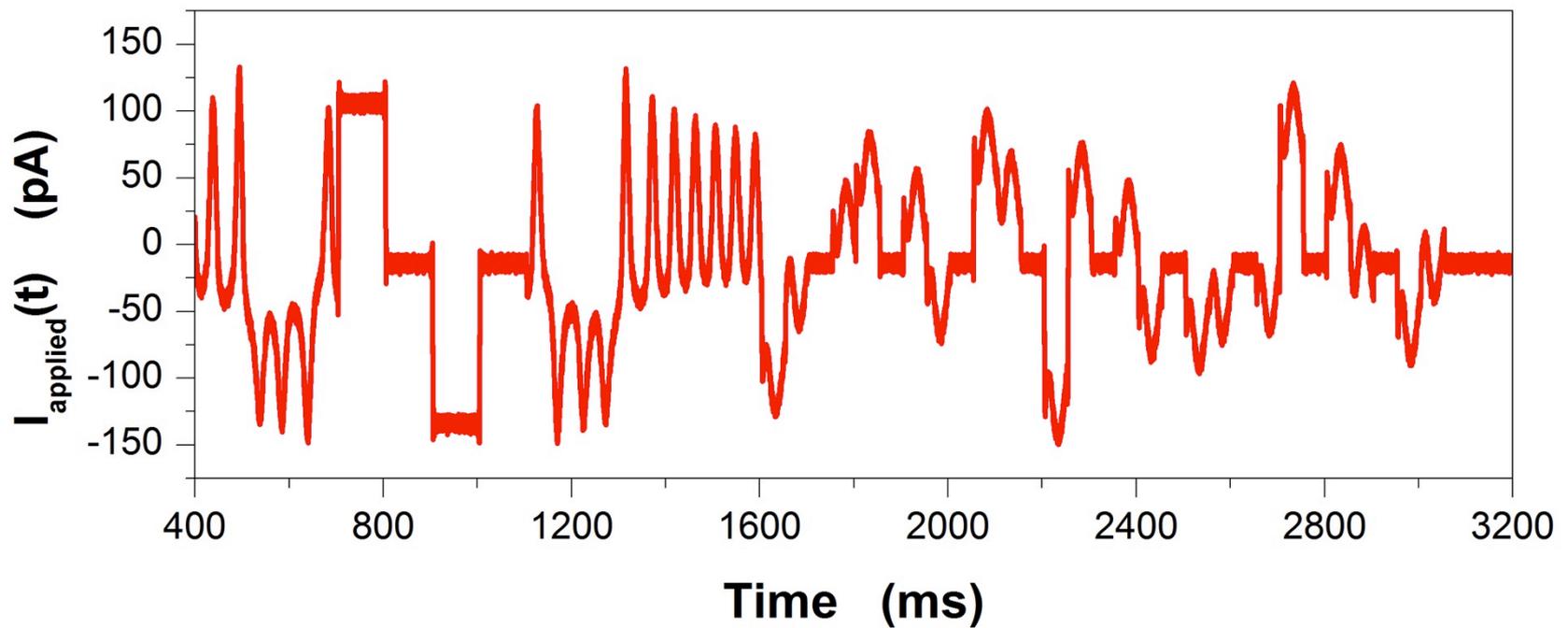
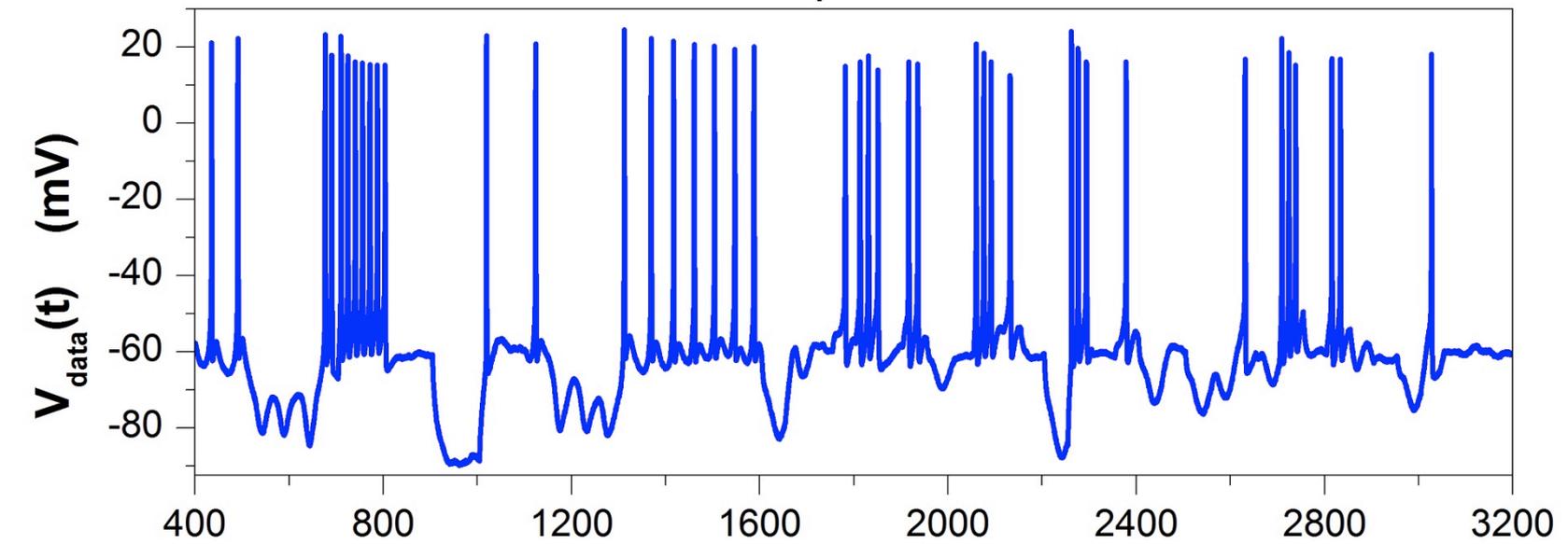
$$\begin{aligned}
C \frac{dV(t)}{dt} = & g_{Na} m(t)^3 h(t) (E_{Na} - V(t)) + \\
& g_K n(t)^4 (E_K - V(t)) + g_L (E_L - V(t)) + \\
& g_{Ca} a(t) b(t) V(t) \frac{[Ca^{2+}]_{ext} - [Ca^{2+}](t) e^{-V(t)/V_T}}{1 - e^{-V(t)/V_T}} \\
& + \text{other currents} + I_{applied}(t)
\end{aligned}$$

$$\frac{dx(t)}{dt} = \frac{x_\infty(V(t) - x(t))}{\tau_x(V(t))} \quad x(t) = \{m(t), h(t), n(t), a(t), b(t)\}$$

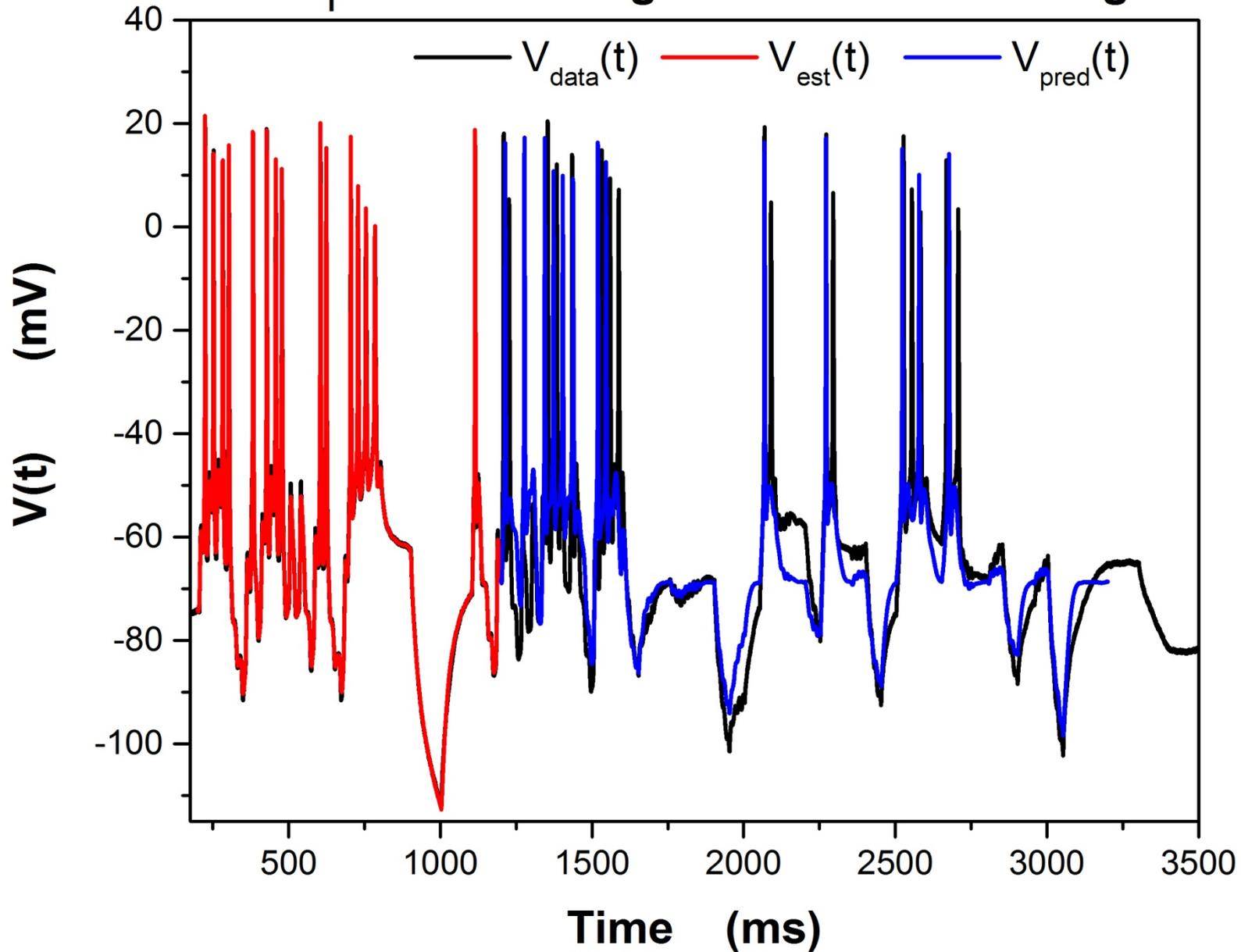
$$x_\infty(V) = \frac{1}{2} \left(1 + \tanh\left(\frac{V - V_x}{dV_x}\right) \right)$$

$$\tau_x(V) = t_{1x} + t_{2x} \left(1 - \tanh^2\left(\frac{V - V_{xt}}{dV_{xt}}\right) \right)$$

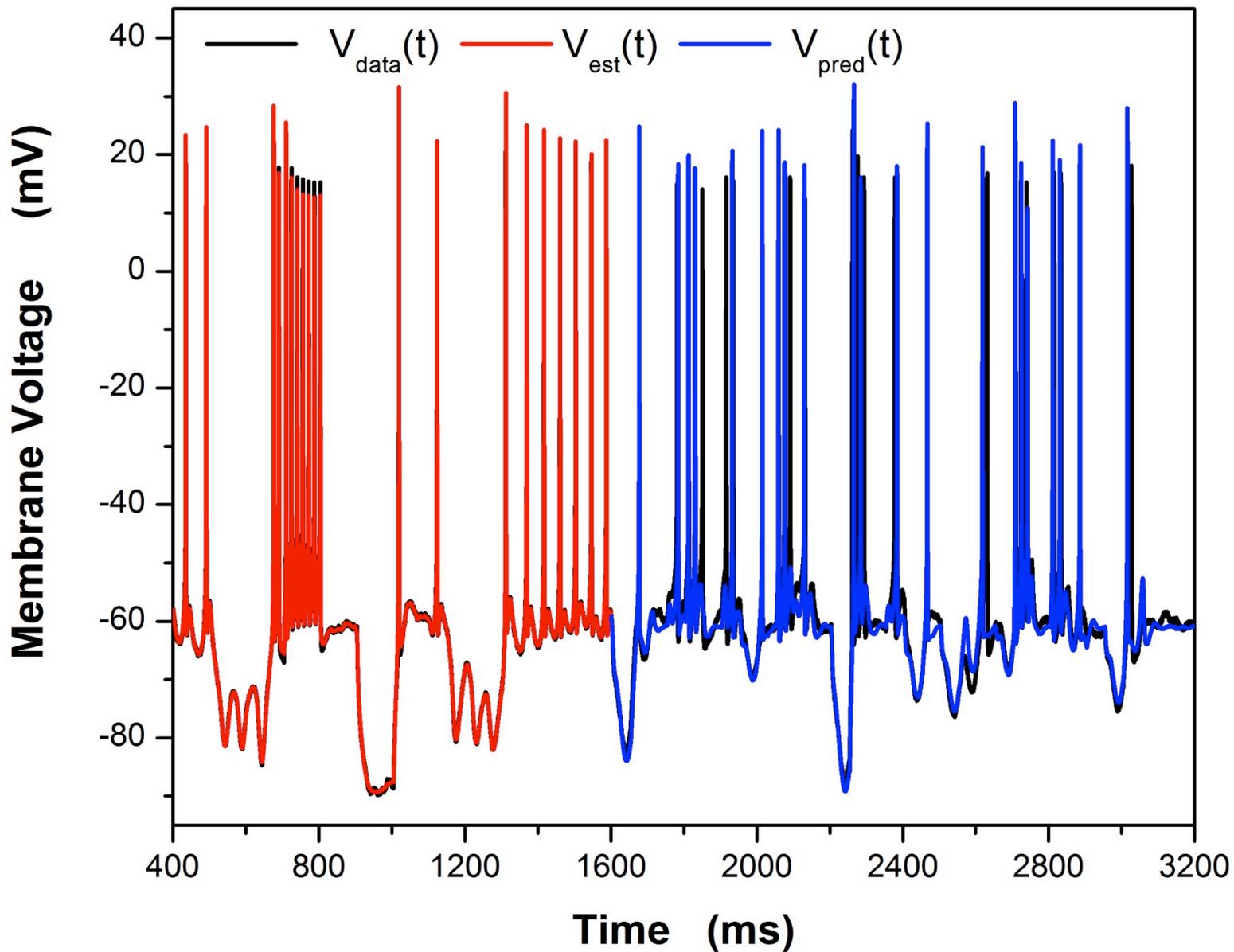
HVC₁ Neuron



HVC₁ Neuron Margoliash/Daou UChicago

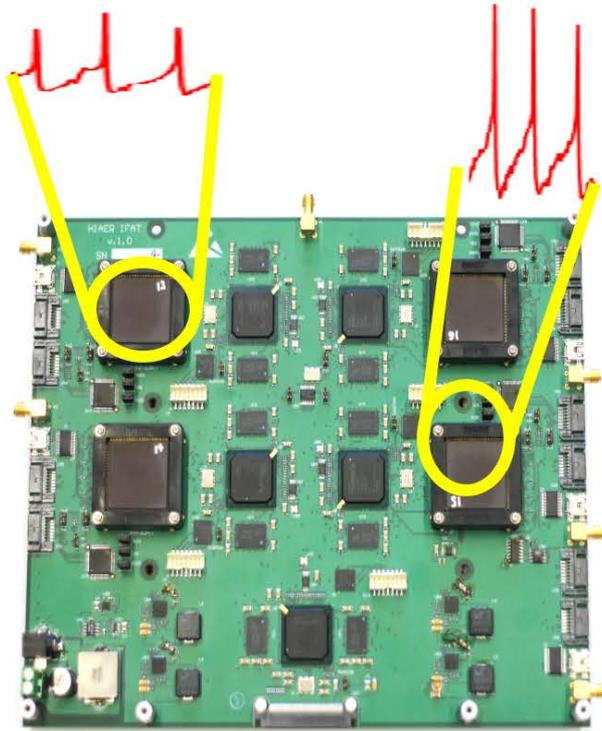


HVC₁ Neuron Data, Estimated, Predicted



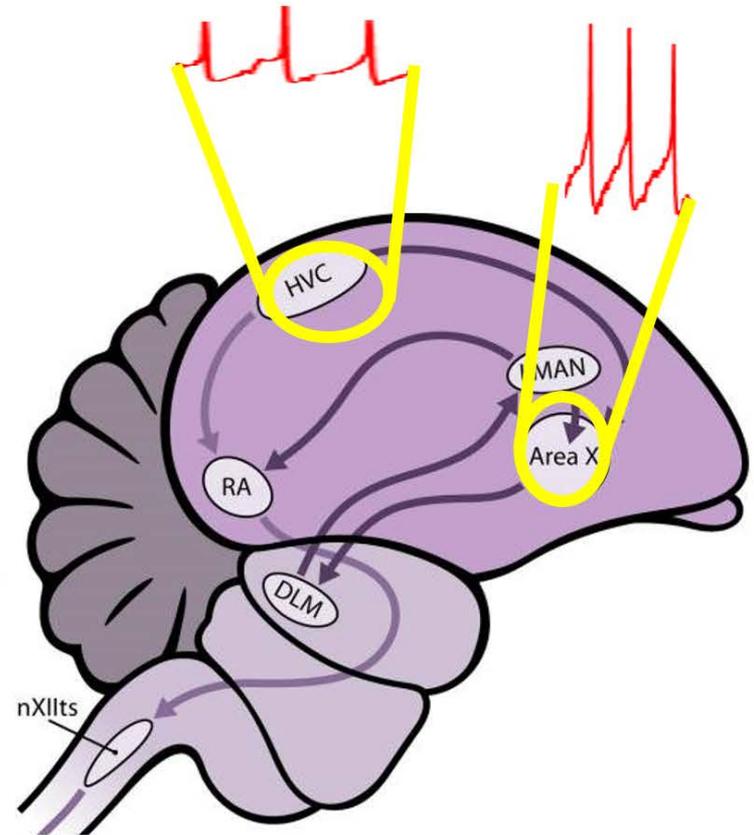
VLSI Neuromorphic Chip

- Test parameters on the chip to check quality of fabrication versus design
- Use a twin experiment to test method of data assimilation: generate data from the VLSI chip. Use “voltages” on chip neurons as measured quantities to estimate parameters known from first step.
- Use voltage data from biological neuron to readjust chip parameters and state variables to those for the data, then predict voltage response to new current stimulation.

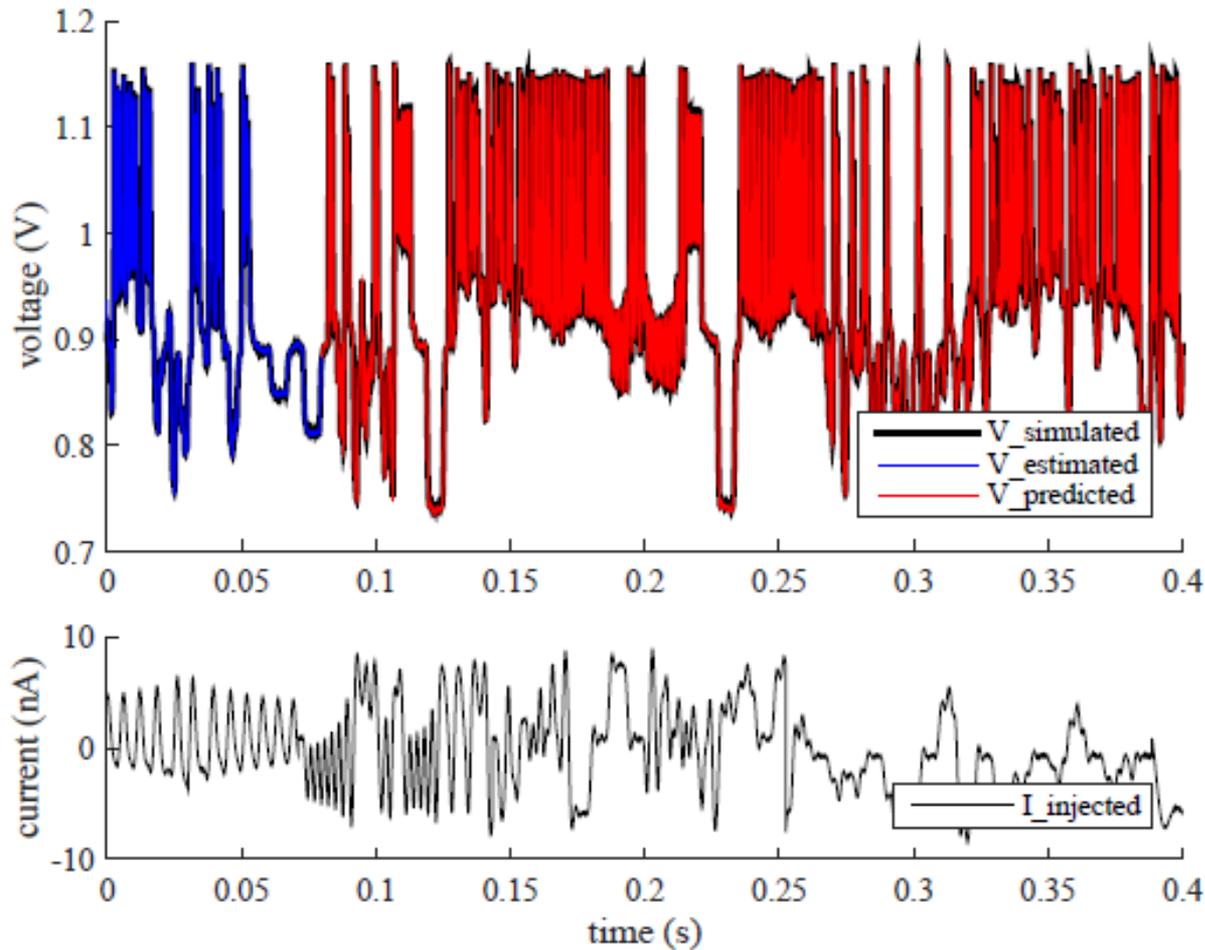


Neuromorphic Chip

**Data
Assimilation**

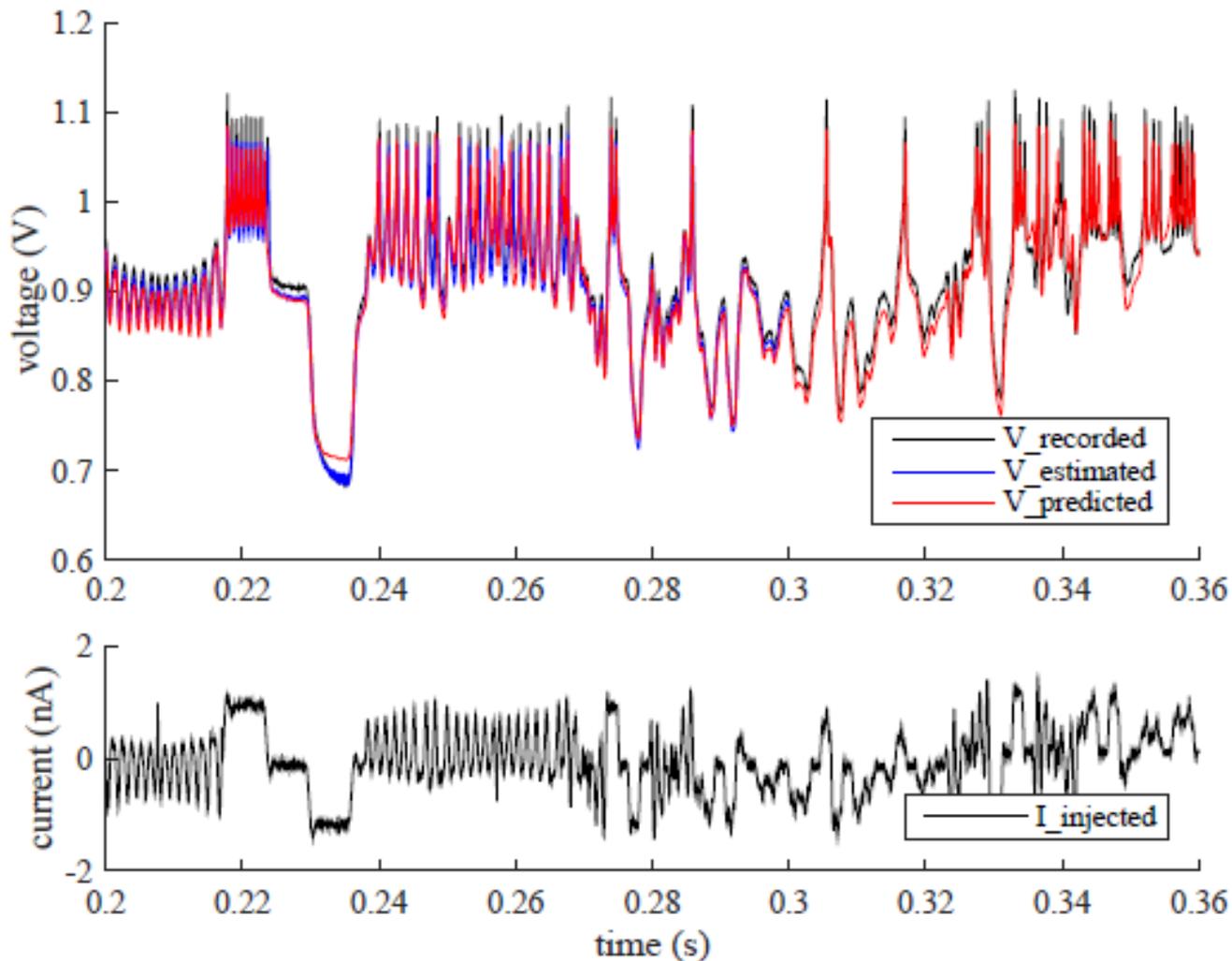


Brain



Test parameters on the chip to check quality of fabrication versus design

Fig. 3. Estimated (blue), predicted (red), and simulated (black) data of the NeuroDyn model in software. The prediction is obtained by integrating the equations of motion forward using the estimated parameters and configuration of the system at the end of the estimation window. The injected stimulating current protocol (black) is also displayed.



Use a twin experiment to test method of data assimilation: generate data from the VLSI chip. Use “voltages” on chip neurons as measured quantities to estimate parameters known from first step.

Fig. 4. Estimated (blue), predicted (red), and measured (black) data recorded from the NeuroDyn chip. The estimation and prediction were obtained, and the current injection was applied, under identical conditions as in Fig. 3.

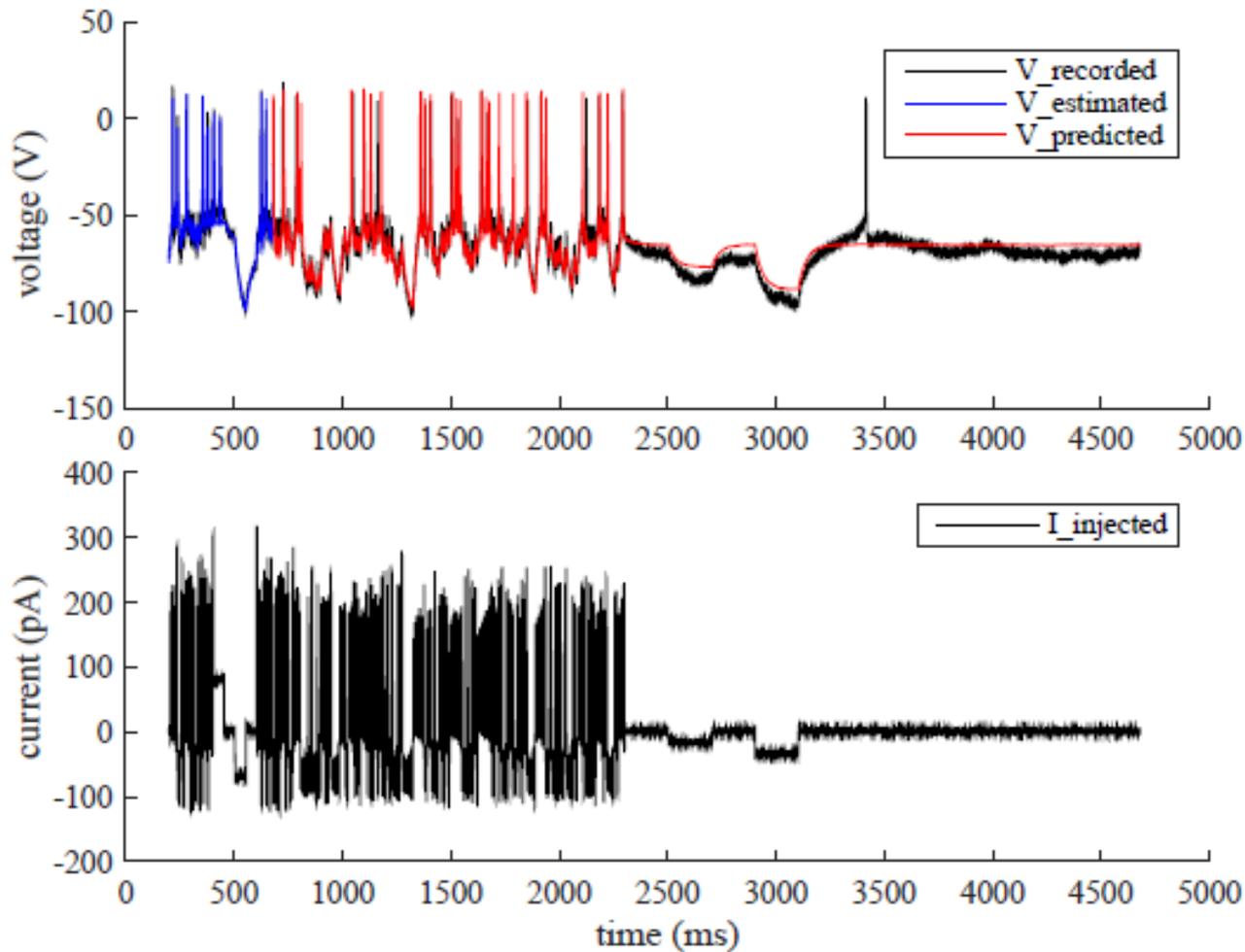


Fig. 5. Estimated (blue), predicted (red), and electrophysiological (black) data recorded *in vivo* from an HVC interneuron (HVC₁) [5] and instantiated onto the NeuroDyn hardware model.

Use voltage data from biological neuron to readjust chip parameters and state variables to those for the data, then predict voltage response to new current stimulation.

So, what did we learn ?

1. Make a model—no algorithms for this, use your best knowledge of the (bio)physics.
2. Make a big model—experiments will prune the model
3. Do twin experiments to determine how many measurements you need to get the “global” minimum of $A_0(X)$ —**annealing**.
4. Use twin experiments to design laboratory experiments
5. Do experiments to determine consistency of model with data.
6. Use the completed model and estimated $x(T)$, via probability distribution or $dx/dt = F(x(t))$, to predict, for $t > T$. This validates (or not) the model.

So, what did we learn ?

7. Use Laplace method + computable corrections to determine consistency of numerical methods.
8. If there are not enough measurements at each observation time, (a) get more; (b) use waveform information via time delays.
9. Using Data Assimilation, one can (a) test new fabrications of VLSI neurons, (b) test DA methods on verified VLSI chip, (c) complete neuron model on chip from biological data; predict response to new forcing.

Unfinished Business

Measurements for Networks of Neurons---extracellular potentials? Other technology

Computational capability for the future

Port network models to VLSI

Use principles of network functions to solve similar problems in other space and time domains.

